



PolyLC INC.

9151 Rumsey Road, Suite 180 • Columbia, MD 21045 USA
Phone: (410) 992-5400 • FAX: (410) 730-8340 • e-mail: POLYLC@aol.com

Instructions for: Capillary and Microbore Columns

Dead Volume:

It is critical to keep this to a minimum. Use connecting tubing of 0.002 - 0.003" i.d. For capillaries < 200 μm i.d., use connecting tubing of 50 μm i.d. Upchurch Scientific sells PEEK tubing in these sizes; an alternative is SGE's PEEKsil material. Keep connections short. This tubing has the following drawbacks:

- 1) Backpressure: A 15-cm length of 0.003" tubing confers @ 500 psi (35 bar) backpressure at a flow rate of 1.0 ml/min.
- 2) Tubing this narrow tends to clog easily. Use inline filters to minimize this tendency.

Detectors:

Flow cells should be no larger than 0.5 μl for microbore columns, and cells for packed capillaries should be 0.1 μl .

Flow Rates:

Assuming a flow rate of 50 $\mu\text{l}/\text{min}$ for a column of 1.0-mm i.d., corresponding flow rates for packed capillaries would be as follows:

Capillary ID	Flow rate	Recommended load or peptides per injection (SCX)	
300 μm	5.0 $\mu\text{l}/\text{min}$	14 μg	While one can load 2x these amounts per run, the resulting peaks may not be as sharp as would be optimal for proteomics applications. Loading levels in HILIC are about 1/4 of those of SCX.
220 μm	2.4 $\mu\text{l}/\text{min}$	7 μg	
175 μm	1.5 $\mu\text{l}/\text{min}$	4 μg	
100 μm	0.5 $\mu\text{l}/\text{min}$	1.4 μg	

Sample Injections:

It is advisable to use either a valve designed for submicroliter sample volumes (*e.g.*, Valco's) or else sample loading via a pressure bomb (@ Lin, Alpert & Yates, *Amer. Genomic/Proteomic Technol.* [Aug. 2001; in press]).

Conditioning, Maintenance, and Storage:

Use the same protocols as with the corresponding columns of conventional dimensions.

Air Bubbles:

It is more difficult to clear these from the detector flow cells than with conventional size columns, owing to the slow flow rates. You can minimize this problem with the following procedures:

- 1) Degas mobile phases.
- 2) Mobile phase should flow from bottom to top into the flow cell.
- 3) Consider installing a flow restrictor after the detector to confer modest additional back pressure.

Death by Mass Spectroscopy:

There is considerable electrical potential at the orifice to a mass spectrometer. The mobile phase can conduct current back to the HPLC system; potentials of 600 V have been measured at injector valves! Be sure to ground the system somewhere between the outlet of the HPLC and the inlet of the MS.